to Laurent⁸ the main component belongs to carbonic anhydrase B. By use of immunoelectrophoretic analyses, with interrupted groove, it was demonstrated that there was a complete reaction of identity between the $3S\gamma_1$ -globulin and the carbonic anhydrase B (Figure 2).

Immunofluorescent technique using a rabbit anti- $3S\gamma_1$ -globulin has revealed that labelling was distributed in the whole cytoplasma of all tubular cells, the glomerule being faintly fluorescent (Figure 3). A $3S\gamma_1$ -antiserum of which the antibodies were precipitated by a pure $3S\gamma_1$ -preparation did not show any further fluorescence.

As we know that carbonic anhydrase is commonly distributed in the kidney, the above data suggest an identity between this enzyme and the $3S\gamma_1$ -globulin. Therefore, we may assume that on the basis of similarities on chemical structure, activity and immunological behaviour, the plasma $3S\gamma$ -globulin and the carbonic anhydrase B are a unique component 9,10 .

Résumé. La $3S\gamma_1$ -globuline, précédemment isolée du plasma humain normal, a fait l'objet d'une comparaison

immunologique avec l'anhydrase carbonique B isolée à partir d'érythrocytes. L'immunoélectrophorèse en gélose et l'immunofluorescence analysée sur le tissu rénal ont montré l'identité immunologique de ces 2 protéines.

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- ⁸ G. LAURENT, M. CHARREL, F. LUCCIONI, M. F. AUTRAN and Y. DERRIEN, Bull. Soc. Chim. biol. 47, 1101 (1965).
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- 10 This work was partially supported by a grant of the Fond National de la Recherche Scientifique.

Detection and Identification of Pyridosine, a Second Lysine Derivative Obtained Upon Acid Hydrolysis of Heated Milk

Recently, a new basic amino acid, found by Erbers-Dobler¹ in an acid hydrolysate of overheated milk, has been identified by Heyns² and Finot³ as ε -N-(2-furoylmethyl)-L-lysine or 'furosine'³. This compound X can also be obtained by acid hydrolysis of ε -N-(1-deoxy-2-ketose)-L-lysine (derivative of glucose (I)³,4; derivative of lactose (II)³).

A second lysine derivative, compound Y, formed simultaneously with furosine during acid hydrolysis of I and II, has now been detected and identified in hydrolysates of overheated milk. This compound cannot be detected by applying the standard conditions of chromatography (Spackman⁵), since it is eluted far after phenylalanine from the 'neutral' column, and well before lysine from the 'basic' column. Its migration is extremely sensitive to changes in pH; the peak can, however, be detected under special conditions (Figures 1 and 2).

Compound Y was isolated by chromatography on Dowex 50 (form H⁺) as a peak eluted after furosine³, and precipitated with methyl ethyl ketone from a methanolic solution of the dried chromatographic fraction. Its structure was determined by UV, IR, NMR and mass spectrometry (operating conditions as previously described³) and confirmed by synthesis.

The mass spectrum of the trifluoroacetyl-methyl ester derivative (molecular peak at m/e 364: $C_{15}H_{19}N_2O_5F_3^+$) shows that compound Y possesses the carbon skeleton of lysine and glucose from which it derives and that only 1 nitrogen can be trifluoroacetylated; the presence of ions at m/e 180, 166, 152, 138, and 124, obtained by elimination of the fragments $(CH_2)_n$ -CH-(NH-CO-CF₃)- CO_2 -CH₃ (n=0, 1, 2, 3, 4) reveals that the α -nitrogen has been trifluoroacetylated; the last 2 fragments (n=3, 4) are formed mostly by rearrangement of 1 hydrogen (m/e = 139, 125) or 2 hydrogens (m/e = 126). The great stability of the ion at m/e 125 indicates that the α -nitrogen belongs to a ring ($C_6H_6NO_2^+$). The ring hypothesis is consistent with the IR bands at 1620 (s), 1550 (s), and 870 (m) cm⁻¹ (γ -pyridone), and is confirmed

by the UV-spectrum (maxima: 277 nm in neutral solution, 300 nm in basic solution, 246 and 275 nm in acidic solution) typical of the 3-hydroxy-4-pyridone structure. The UV-spectrum is similar to that of a natural compound, β -(1, 4-dihydro-3-hydroxy-4-oxo-1-pyridyl)-alanine, namely mimosine or leucenol (maxima: 280 nm in neutral solution, 305 nm in basic solution, and 272 with shoulder at 250 nm in acidic solution) isolated by Renz⁶ and synthesized by Adams⁷ and Spencer⁸.

The remaining substituents of the ring are given by the NMR-spectrum (Figure 3): singlets at δ (ppm) 8.16 and 7.20 (2 non coupled hydrogens), and at 2.65 (methyl). The methyl group must be either in position 5 or 6; the more probable position 6 has been confirmed by synthesis. The latter has been performed using the technique of Heyns⁹, by condensing α -N-formyl-L-lysine with the corresponding γ -pyrone, allomaltol, prepared according to Brown ¹⁰; the reference product, namely ε -(1, 4-dihydro-6-methyl-3-hydroxy-4-oxo-1-pyridyl)-L-lysine (V), has the same spectroscopic behaviour as compound Y. The trivial name pyridosine is proposed for this new amino acid.

- ¹ H. Erbersdobler and H. Zucker, Milchwissenschaft 21, 564 (1966).
- ² K. Heyns, J. Heukeshoven and K. H. Brose, Angew. Chem. 80, 627 (1968).
- ³ P. A. FINOT, J. BRICOUT, R. VIANI and J. MAURON, Experientia 24, 1097 (1968).
- ⁴ J. Bruggemann and H. Erbersdobler, Z. Lebensmittelunters. u. -Forsch. 137, 137 (1968).
- ⁵ D. H. SPACKMAN, W. H. STEIN and S. MOORE, Analyt. Chem. 30, 1185 (1958).
- ⁶ J. Renz, Hoppe-Seylers Z. physiol. Chem. 244, 153 (1936).
- ⁷ R. Adams and J. L. Johnson, J. Am. chem. Soc. 71, 705 (1949).
- I. D. Spencer and A. D. Notation, Can. J. Chem. 40, 1374 (1962).
 K. Heyns and G. Vogelsang, Chem. Ber. 87, 1377 (1954).
- ¹⁰ M. G. Brown, J. chem. Soc. 2558 (1956).

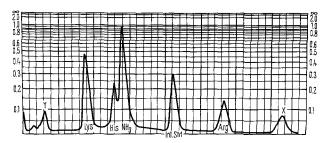


Fig. 1. Basic amino acid chromatography of a scorched roller dried milk after 6N HCl hydrolysis. X, furosine; Y, second hydrolysis product of inactivated lysine; internal standard (Int Std), 2-amino-granidino-propionic acid. Conditions: Resin PA-35 Beckman; column height, 9.5 cm; diam. 0.9 cm; buffer $0.38\,N$ pH 4.26: 16 ml; then buffer $0.35\,N$ pH 5.28; speed: 50 ml/h.

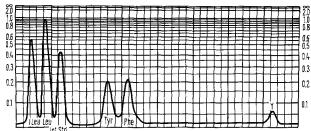


Fig. 2. Neutral amino acid chromatography of a scorched roller dried milk after 6N HCl hydrolysis. Y, second hydrolysis product of inactivated lysine. Conditions: Resin PA-35 Beckman; column height, 50 cm; diam. 0.9 cm; buffer 0.2N pH 4.25 until the complete elution of the internal standard peak (norleucine); then buffer 0.35N pH 5.28; speed: 50 ml/h.

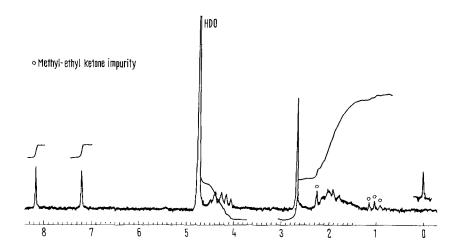


Fig. 3. NMR-spectrum of compound Y in D_2O . Internal standard: sodium 3-(trimethylsilyl)-propansulfonate (Merck, Darmstadt).

This isomer of furosine probably derives from the β -furanose form of I and II, through III, which in acidic solution can easily yield IV (ELMING¹¹)

The equilibrium between IV (aromatic cation) and V (neutral molecule) explains the delayed migration of compound Y on polystyrene sulfonated resins used with acidic buffers, in spite of its neutral behaviour on paper electrophoresis at pH 3.9 according to BISERTE 12.

In acid hydrolysates of treated milk, the determination of the 2 products obtained from inactivated lysine

(furosine and pyridosine) could be a quick checking of the loss in available lysine provoked by the Maillard reaction. A systematic investigation of the hydrolysates of lysine inactivated by reducing sugars, might then enlarge the field of application of this technique to any foodstuff¹³.

Résumé. Un deuxième dérivé de la lysine inactivée, l'ε-(1,4-dihydro-6-methyl-3-hydroxy-4-oxo-1-pyridyl)-L-lysine, a été trouvé dans les hydrolysats de laits surchauffés; il se forme à partir de la deoxy-D-lactulose-L-lysine et peut être mis en évidence dans des conditions particulières de chromatographie. Sa structure a été déterminée par spectrométrie de masse, UV, IR et RMN; le nom de pyridosine est proposé pour ce nouvel acide aminé.

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¹¹ N. Elming and N. Clauson-Kaas, Acta chem. scand. 10, 1603 (1956).

¹² G. BISERTE, T. PLAQUET-SCHOONAERT, P. BOULANGER and P. PAY-SANT, J. Chromat. 3, 25 (1960).

¹³ We wish to thank Prof. H. Prinzbach of the University of Lausanne for the NMR-spectrum and Miss E. Bujard for the amino acid analysis by ion exchange chromatography.